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     FILE 'HCAPLUS' ENTERED AT 15:59:57 ON 25 NOV 2008
L1       489325 S RNA OR (NUCLEIC ACID)
L2      1519165 S ALKALI OR LITHIUM OR SALT
L3       168678 S DETERGENT OR AMPHIPHILIC OR TRITON OR TWEEN
L4          351 S L1 AND L2 AND L3
L5          216 S L4 AND (PY<2001 OR AY<2001 OR PRY<2001)

     FILE 'STNGUIDE' ENTERED AT 16:02:12 ON 25 NOV 2008

     FILE 'HCAPLUS' ENTERED AT 16:02:29 ON 25 NOV 2008
L6      1168343 S SOLID
L7           21 S L5 AND L6
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=> file hcaplus
COST IN U.S. DOLLARS                                SINCE FILE      TOTAL
                                                       ENTRY    SESSION
FULL ESTIMATED COST                                     0.21       0.21

FILE 'HCAPLUS' ENTERED AT 15:59:57 ON 25 NOV 2008
USE IS SUBJECT TO THE TERMS OF YOUR STN CUSTOMER AGREEMENT.
PLEASE SEE "HELP USAGETERMS" FOR DETAILS.
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FILE COVERS 1907 - 25 Nov 2008  VOL 149 ISS 22
FILE LAST UPDATED: 24 Nov 2008  (20081124/ED)

HCAplus now includes complete International Patent Classification (IPC)
reclassification data for the third quarter of 2008.

New CAS Information Use Policies, enter HELP USAGETERMS for details.

 This file contains CAS Registry Numbers for easy and accurate
 substance identification.

=> s RNA or (nucleic acid)
        361046 RNA
        220862 NUCLEIC
       4714979 ACID
        167978 NUCLEIC ACID
                 (NUCLEIC(W)ACID)
L1      489325 RNA OR (NUCLEIC ACID)

=> s alkali or lithium or salt
        433788 ALKALI
        351400 LITHIUM
        867025 SALT
L2     1519165 ALKALI OR LITHIUM OR SALT

=> s detergent or amphiphilic or triton or tween
         86591 DETERGENT
         21703 AMPHIPHILIC
         45671 TRITON
         24812 TWEEN
L3      168678 DETERGENT OR AMPHIPHILIC OR TRITON OR TWEEN

=> s l1 and l2 and l3
L4         351 L1 AND L2 AND L3

=> s l4 and (PY<2001 or AY<2001 or PRY<2001)
      21007117 PY<2001
       3943599 AY<2001
       3412660 PRY<2001
L5         216 L4 AND (PY<2001 OR AY<2001 OR PRY<2001)
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=> file stnguide
COST IN U.S. DOLLARS                                SINCE FILE      TOTAL
                                                       ENTRY    SESSION
FULL ESTIMATED COST                                    10.76      10.97

FILE 'STNGUIDE' ENTERED AT 16:02:12 ON 25 NOV 2008
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FILE CONTAINS CURRENT INFORMATION.
LAST RELOADED: Nov 21, 2008 (20081121/UP).

=> file hcaplus
COST IN U.S. DOLLARS                                SINCE FILE      TOTAL
                                                       ENTRY    SESSION
FULL ESTIMATED COST                                     0.06      11.03

FILE 'HCAPLUS' ENTERED AT 16:02:29 ON 25 NOV 2008
USE IS SUBJECT TO THE TERMS OF YOUR STN CUSTOMER AGREEMENT.
PLEASE SEE "HELP USAGETERMS" FOR DETAILS.
COPYRIGHT (C) 2008 AMERICAN CHEMICAL SOCIETY (ACS)
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FILE COVERS 1907 - 25 Nov 2008  VOL 149 ISS 22
FILE LAST UPDATED: 24 Nov 2008  (20081124/ED)

HCAplus now includes complete International Patent Classification (IPC)
reclassification data for the third quarter of 2008.

New CAS Information Use Policies, enter HELP USAGETERMS for details.

 This file contains CAS Registry Numbers for easy and accurate
 substance identification.

=> s solid
L6     1168343 SOLID

=> s l5 and l6
L7           21 L5 AND L6

=> d l7 1-21 ti abs bib

L7   ANSWER 1 OF 21  HCAPLUS  COPYRIGHT 2008 ACS on STN
TI   Compositions, methods, and kits for isolating nucleic acids using
     surfactants and proteases
AB   The invention relates to compns. and methods for isolating nucleic acids
     from biol. samples, including whole tissue.  The invention also provides
     kits for isolating nucleic acids from biol. samples.  A method for
     obtaining nucleic acid from a biol. sample and binding
     the nucleic acid to a solid phase comprises
```

(a) contacting the biol. sample with a disrupting buffer, wherein the disrupting buffer comprises a protease and a cationic surfactant; (b) substantially neutralizing the cationic surfactant; and (c) binding the nucleic acid to a solid phase. Genomic DNA was isolated from several rat tissues and mouse tail using a digestion solution containing 1 mg of Proteinase K, 1 % DTAB, 100 mM Tris-HCl (pH 8.0), 20 μM ATA, and 20 mM CaCl2 and incubating for 60 min at 65°. Most of the tissues were effectively digested in less than one hour. Digestion of liver, brain and kidney were about 95 % complete after one hour. Following digestion, binding solution containing 5 M GuSCN, 50 mM MES (pH 6.0), 20 mM EDTA, and 6 % Tween 20 was then added to each sample and the samples were placed on GF/B filter membranes for washing and recovery of DNA.

AN 2002:907069 HCAPLUS <<LOGINID::20081125>>
DN 138:1959
TI Compositions, methods, and kits for isolating nucleic acids using surfactants and proteases
IN Greenfield, Lawrence; Montesclaros, Luz
PA Applera Corp., USA
SO U.S. Pat. Appl. Publ., 57 pp., Cont.-in-part of U.S. Ser. No. 724,613.
CODEN: USXXCO
DT Patent
LA English
FAN.CNT 2

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE	
PI	US 20020177139	A1	20021128	US 2001-997169	20011128	<--
	US 6762027	B2	20040713			
	US 7001724	B1	20060221	US 2000-724613	20001128	<--
	US 20050009045	A1	20050113	US 2004-800137	20040311	<--
	US 7303876	B2	20071204			
	JP 2006197941	A	20060803	JP 2006-74844	20060317	<--
PRAI	US 2000-724613	A2	20001128	<--		
	JP 2002-587600	A3	20011128			
	US 2001-997169	A1	20011128			

RE.CNT 28 THERE ARE 28 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L7 ANSWER 2 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Methods for decreasing non-specific binding of beads in dual bead assays including related optical biodiscs and disc drive systems
AB Methods are disclosed for decreasing non-specific bindings of beads in dual bead assays and related optical bio-disks and disk drive systems. The methods are employed to determine the suitability of a test solid phase for use in a dual bead assay. The methods include identifying whether a target agent is present in a biol. sample and involve mixing capture beads, each having at least one transport probe affixed thereto. Reporter beads each have at least one signal probe affixed thereto. The reporter and capture beads are each bound to the target agent. The methods further include isolating the dual bead complex from the mixture to obtain an isolate, and exposing the isolate to a capture field on a disk. Detecting the presence of the dual bead complex in the disk is then performed to determine whether the target agent is present in the sample. The method further includes pre-treating capture beads and reporter beads with detergents prior to capture, treating capture beads and reporter beads with blocking agents prior to target capture, and performing the mixing in an intermittent manner. The beads are preferably mixed only when they start to settle down in the tube or on the disk. The methods also provide for evaluation of non-specific binding of the dual bead assay in the presence of salt concns. ranging from 0.1M up to 1M and use of a

new wash buffer having 10 mM EDTA. Dual bead DNA hybridization assays were made.

AN 2002:889450 HCAPLUS <<LOGINID::20081125>>

DN 137:365966

TI Methods for decreasing non-specific binding of beads in dual bead assays including related optical biodiscs and disc drive systems

IN Phan, Brigitte Chau; Lam, Amethyst Hoang; Yeung, Kayuen

PA USA

SO U.S. Pat. Appl. Publ., 77 pp., Cont.-in-part of U.S. Ser. No. 997,741.
CODEN: USXXCO

DT Patent

LA English

FAN.CNT 28

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 20020172980	A1	20021121	US 2002-87549	20020228 <--
	US 20030003464	A1	20030102	US 2001-997741	20011127 <--
	WO 2002071929	A2	20020919	WO 2002-US7955	20020314
	WO 2002071929	A3	20030320		
	W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW				
	RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG				
	WO 2002073605	A2	20020919	WO 2002-US8208	20020314
	WO 2002073605	A3	20030403		
	W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZM, ZW				
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	AU 2002258528	A1	20020924	AU 2002-258528	20020314
	AU 2002306750	A1	20020924	AU 2002-306750	20020314
PRAI	US 2000-253283P	P	20001127	<--	
	US 2000-253958P	P	20001128	<--	
	US 2001-272134P	P	20010228		
	US 2001-272243P	P	20010228		
	US 2001-272525P	P	20010301		
	US 2001-275006P	P	20010312		
	US 2001-275643P	P	20010314		
	US 2001-278691P	P	20010326		
	US 2001-314906P	P	20010824		
	US 2001-997741	A2	20011127		
	US 2002-352270P	P	20020130		
	US 2001-278106P	P	20010323		
	US 2001-278110P	P	20010323		
	US 2001-278688P	P	20010326		
	US 2001-278694P	P	20010326		
	US 2001-278697P	P	20010326		
	US 2001-911253	A	20010723		
	US 2002-38297	A	20020104		

	WO 2002-US7955	W	20020314		
	WO 2002-US8208	W	20020314		

L7 ANSWER 3 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Isolation of nucleic acids from biological samples using surfactants and proteases
AB The invention relates to compns. and methods for isolating nucleic acids from biol. samples, including whole tissue. The method comprises contacting the biol. sample with a disrupting buffer containing proteases (e.g., Proteinase K) and a cationic surfactant (e.g., CTAB). The cationic surfactant is then neutralized either by its removal or by use of a second nonionic surfactants (e.g., Tween 20). Nucleic acids are then isolated by binding to a solid phase, such as glass fiber GF/B filters. The effects of cationic surfactants on activity of proteinase K, and the solubility of surfactants in different chaotropes is investigated to identify optimal cationic surfactants and salts. The invention also provides kits for isolating nucleic acids from biol. samples.
AN 2002:869079 HCAPLUS <<LOGINID::20081125>>
DN 137:365972
TI Isolation of nucleic acids from biological samples using surfactants and proteases
IN Greenfeld, I. Larry
PA PE Corporation, USA; Applera Corporation
SO PCT Int. Appl., 129 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 2

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 2002090539	A2	20021114	WO 2001-US45071	20011128 <--
	WO 2002090539	A3	20030807		
	W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZM, ZW				
	RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG				
	US 7001724	B1	20060221	US 2000-724613	20001128 <--
	CA 2429941	A1	20021114	CA 2001-2429941	20011128 <--
	AU 2001297835	A1	20021118	AU 2001-297835	20011128 <--
	AU 2001297835	B2	20051110		
	EP 1354036	A2	20031022	EP 2001-274041	20011128 <--
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, MK, CY, AL, TR				
	JP 2005501523	T	20050120	JP 2002-587600	20011128 <--
	JP 2006197941	A	20060803	JP 2006-74844	20060317 <--
PRAI	US 2000-724613	A	20001128	<--	
	JP 2002-587600	A3	20011128		
	WO 2001-US45071	W	20011128		

L7 ANSWER 4 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Methods and kits for isolating nucleic acids from leukocytes by binding to antibodies on a solid support
AB The present invention relates to a method of isolating nucleic acid from a blood sample. The method involves selectively isolating leukocytes from said sample by binding said leukocytes to a

solid support containing a binding partner specific for the leukocyte, for example an antibody. The antibody can bind an antigen selected from one of more of the following: HLA-I, CD11a, CD18, CD45, CD46, CD50, CD82, CD162, CD5 and CD15 and a specific example shows a combination of CD45 and CD15. The said leukocytes are lysed in detergents to release nucleic acids which are subsequently bound to a second solid support which is neg. charged. Kits for isolating nucleic acid from samples form further embodiments of the invention.

AN 2001:904506 HCAPLUS <<LOGINID::20081125>>
DN 136:15912
TI Methods and kits for isolating nucleic acids from leukocytes by binding to antibodies on a solid support
IN Bergholtz, Stine; Korsnes, Lars; Andreassen, Jack
PA Dynal Biotech Asa, Norway; Jones, Elizabeth Louise
SO PCT Int. Appl., 51 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI WO 2001094572	A1	20011213	WO 2001-GB2472	20010605 <--
W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW				
RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG				
CA 2410888	A1	20011213	CA 2001-2410888	20010605 <--
CA 2410888	C	20080916		
EP 1290155	A1	20030312	EP 2001-934205	20010605 <--
EP 1290155	B1	20060809		
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, MK, CY, AL, TR				
AU 2001260507	B2	20060831	AU 2001-260507	20010605 <--
AT 335815	T	20060915	AT 2001-934205	20010605 <--
ES 2269399	T3	20070401	ES 2001-934205	20010605 <--
US 20030180754	A1	20030925	US 2003-297301	20030430 <--
PRAI GB 2000-13658	A	20000605	<--	
WO 2001-GB2472	W	20010605		

RE.CNT 4 THERE ARE 4 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L7 ANSWER 5 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Method for extracting nucleic acid
AB A simple method is provided for easily extracting nucleic acid (e.g., DNA) even from a sample containing impurities (e.g, blood) without using a substance harmful to an environment or a human body. The method comprises a process of mixing the sample containing nucleic acid with a dissolving solution containing a surfactant, a salt , a buffer, and a chelating agent, a process for contacting the mixture solution with a hydrophilic surface-possessing solid phase carrier support consisting of a polymer possessing phosphate ester portions at least as a part of structural unit, and a process for isolating the solid phase support from the dissolving solution
AN 2001:551708 HCAPLUS <<LOGINID::20081125>>
DN 135:133096
TI Method for extracting nucleic acid

IN Kawamura, Kiyoko; Kitahiro, Tsuneji; Oshima, Kunihiro; Yamamoto, Ryohei
PA Kurashiki Spinning Co., Ltd., Japan
SO Jpn. Kokai Tokkyo Koho, 13 pp.
    CODEN: JKXXAF
DT Patent
LA Japanese
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	JP 2001204462	A	20010731	JP 2000-14391	20000124 <--
	JP 3397737	B2	20030421		
PRAI	JP 2000-14391		20000124	<--	

L7 ANSWER 6 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Templating of solid particles by polymer multilayers
AB The invention is directed to (i) the encapsulation of elec. uncharged organic substances in polymeric capsules by using a multi-step strategy that involves charging the surface of the microcrystals with an amphiphilic substance, followed by consecutively depositing polyelectrolytes of opposite charge to assemble a multilayered shell of polymeric material around the microcrystal template, and (ii) the formation of polymer multilayer cages derived from the coated crystals by facile removal of the crystalline template.
AN 2001:524661 HCAPLUS <<LOGINID::20081125>>
DN 135:108256
TI Templating of solid particles by polymer multilayers
IN Caruso, Frank; Mohwald, Helmuth; Trau, Dieter; Renneberg, Reinhard
PA Max-Planck-Gesellschaft Zur Forderung Der Wissenschaften E.V., Germany
SO Eur. Pat. Appl., 23 pp.
    CODEN: EPXXDW
DT Patent
LA English
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	EP 1116516	A1	20010718	EP 2000-111523	20000529 <--
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO				
	DE 10001172	A1	20010726	DE 2000-10001172	20000113 <--
	WO 2001051196	A1	20010719	WO 2001-EP329	20010112 <--
	W: JP, US				
	RW: AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR				
	EP 1246692	A1	20021009	EP 2001-903643	20010112 <--
	EP 1246692	B1	20050330		
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI, CY, TR				
	JP 2003519565	T	20030624	JP 2001-551606	20010112 <--
	AT 291958	T	20050415	AT 2001-903643	20010112 <--
	ES 2236175	T3	20050716	ES 2001-903643	20010112 <--
	US 20020187197	A1	20021212	US 2002-148890	20020617 <--
	US 7045146	B2	20060516		
PRAI	DE 2000-10001172	A	20000113	<--	
	EP 2000-111523	A	20000529	<--	
	WO 2001-EP329	W	20010112		

RE.CNT 6 THERE ARE 6 CITED REFERENCES AVAILABLE FOR THIS RECORD
           ALL CITATIONS AVAILABLE IN THE RE FORMAT

L7 ANSWER 7 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Apparatus for conducting chemical or biochemical reactions on a solid surface within an enclosed chamber

AB The invention provides an apparatus and method for conducting chemical or biochem.
reactions on a solid surface within an enclosed chamber. The invention may be used in conducting hybridization reactions, as of biopolymers such as DNA, RNA, oligonucleotides, peptides, polypeptides, proteins, antibodies, and the like. In another aspect, the invention provides an improved method for mixing a thin film of solution, as in a hybridization chamber. The invention further provides a kit for carrying out the methods of the invention. In a nucleic acid hybridization assay, background interference was low when hybridization solution containing 1 weight% Triton X-100 was used.

AN 2001:499793 HCAPLUS <<LOGINID::20081125>>

DN 135:89490

TI Apparatus for conducting chemical or biochemical reactions on a solid surface within an enclosed chamber

IN Schembri, Carol T.; Overman, Leslie B.; Hotz, Charles Z.

PA Agilent Technologies Inc., USA

SO U.S., 17 pp.
CODEN: USXXAM

DT Patent

LA English

FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 6258593	B1	20010710	US 1999-343372	19990630 <--
	US 20020001839	A1	20020103	US 2001-900294	20010706 <--
	US 6911343	B2	20050628		
	US 20050250129	A1	20051110	US 2005-41129	20050121 <--
	US 7247499	B2	20070724		
PRAI	US 1999-343372	XX	19990630	<--	
	US 2001-900294	A1	20010706		

RE.CNT 31 THERE ARE 31 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L7 ANSWER 8 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN

TI Reverse transcriptase assay kit, its diagnostic use and method for analysis of RT activity in biological samples

AB A reverse transcriptase (RT) assay kit for anal. of RT activity in biol. samples is described. The kit comprises solid phase bound poly(rA) and/or poly(dA) template(s) obtainable by contacting a polystyrene-based solid phase with a 1-methylimidazole-containing coupling solution, and RT-type adapted assay components selected from a buffer, divalent metal ion, chelator, polyamine, RNase inhibitor, reducing agent, salt, stabilizing agent, and detergent, and deoxynucleotide triphosphate, primer, protective agent and concentrated washing buffer, and optionally lyophilized reference enzyme(s), and further optionally lyophilized alkaline phosphatase conjugated anti-BrdU monoclonal antibody, alkaline phosphatase substrate buffer and alkaline phosphatase substrate, and written instructions for use of the assay kit. Further, a method and a use of the assay kit for the qual. and quant. anal. of RT activity in a biol. sample, optionally followed by evaluation of the status of a RT activity related disorder or disease based on the result of the anal. of the RT activity, are disclosed.

AN 2001:12719 HCAPLUS <<LOGINID::20081125>>

DN 134:53129

TI Reverse transcriptase assay kit, its diagnostic use and method for analysis of RT activity in biological samples

IN Kallander, Clas; Gronowitz, Simon; Pettersson, Ingvar

PA Cavidi Tech AB, Swed.

SO PCT Int. Appl., 32 pp.
CODEN: PIXXD2

DT Patent
LA English
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE	
PI	WO 2001001129	A2	20010104	WO 2000-EP5563	20000616	<--
	WO 2001001129	A3	20010726			
	W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW					
	RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG					
	AU 2000061503	A	20010131	AU 2000-61503	20000616	<--
	AU 764566	B2	20030821			
	EP 1185705	A2	20020313	EP 2000-947849	20000616	<--
	EP 1185705	B1	20040818			
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO					
	BR 2000011812	A	20020402	BR 2000-11812	20000616	<--
	JP 2003503073	T	20030128	JP 2001-507084	20000616	<--
	AT 274070	T	20040915	AT 2000-947849	20000616	<--
	PT 1185705	T	20041231	PT 2000-947849	20000616	<--
	ES 2225179	T3	20050316	ES 2000-947849	20000616	<--
	CN 1218047	C	20050907	CN 2000-809387	20000616	<--
	ZA 2001010064	A	20021206	ZA 2001-10064	20011206	<--
	IN 2001MN01545	A	20070622	IN 2001-MN1545	20011206	<--
	MX 2001PA12951	A	20030624	MX 2001-PA12951	20011214	<--
	US 6849406	B1	20050201	US 2001-926808	20011221	<--
	IN 2005MN00299	A	20051202	IN 2005-MN299	20050418	<--
PRAI	SE 1999-2410	A	19990624	<--		
	WO 2000-EP5563	W	20000616	<--		
	IN 2001-MN1545	A3	20011206			

L7 ANSWER 9 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN

TI Rapid and efficient method for isolating nucleic acids from any complex starting material without use of chaotropic salts

AB The invention relates to formulations without chaotropic components for isolating nucleic acids, notably DNA, from any quantity of any complex starting material, by bonding to a solid phase. The formulations contain a lysis/bonding buffer system presenting at least one antichaotropic salt component such as ammonium chloride, a solid phase, and known washing and elution buffers. The lysis/bonding buffer system can be an aqueous solution or a solid formulation in ready-to-use reaction vessels. As the solid phase any support materials are suitable which are used for isolation by means of chaotropic reagents, such as, preferably, glass-fiber matting, glass membranes, silicon supports, ceramic materials, zeolites, or materials having neg. functionalized surfaces or chemical modified surfaces which can be given a neg. charge potential. The invention further relates to a method for isolating nucleic acids, notably DNA, from any complex starting materials by using the formulations provided for in the invention. Said method is characterized by the following: lysis of the starting material, bonding of the nucleic acids to a support material, washing of the nucleic acids bound to said support, and elution of the nucleic acids.

AN 2000:401988 HCAPLUS <<LOGINID::20081125>>

DN 133:27347

TI Rapid and efficient method for isolating nucleic acids from any complex starting material without use of chaotropic salts
IN Hillebrand, Timo; Bendzko, Peter
PA Invitek G.m.b.H., Germany
SO PCT Int. Appl., 49 pp.
CODEN: PIXXD2
DT Patent
LA German
FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI WO 2000034463	A1	20000615	WO 1999-DE2248	19990723 <--
W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW				
RW: GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG				
DE 19856064	A1	20000629	DE 1998-19856064	19981204 <--
DE 19856064	C2	20001130		
CA 2352472	A1	20000615	CA 1999-2352472	19990723 <--
AU 9961862	A	20000626	AU 1999-61862	19990723 <--
EP 1135479	A1	20010926	EP 1999-948662	19990723 <--
EP 1135479	B1	20021218		
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO				
JP 2002531126	T	20020924	JP 2000-586897	19990723 <--
AT 230022	T	20030115	AT 1999-948662	19990723 <--
RU 2241004	C2	20041127	RU 2001-118287	19990723 <--
CN 1277922	C	20061004	CN 1999-815132	19990723 <--
US 20010041332	A1	20011115	US 1999-454740	19991206 <--
US 6699987	B2	20040302		
NO 2001002700	A	20010730	NO 2001-2700	20010601 <--
PRAI DE 1998-19856064	A	19981204 <--		
WO 1999-DE2248	W	19990723 <--		

RE.CNT 7 THERE ARE 7 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L7 ANSWER 10 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Methods for detecting and measuring spliced nucleic acids and method of cytoplasmic nucleic acid preparation
AB A simplified method for preparing a biol. sample to release cytoplasmic nucleic acid, preferably spliced mRNA, suitable for amplification, while minimizing the release of nuclear genetic material is disclosed. A buffer containing a soluble salt with ionic strength of particular range and a non-ionic detergent are used to lyse the cells. MRNA is then purified by contacting the sample with a solid support joined to an immobilized oligonucleotide which would form stable hybridization complex with the mRNA. Immobilized oligonucleotide preferably contains a poly-T sequence. A method of detecting and measuring the amount of fusion nucleic acid , notably spliced mRNA present in the sample, following nucleic acid amplification, is also disclosed. A fusion nucleic acid to be detected contain a splice junction site, and primers designed to have sequences complementary to and around the splice-junction site are used to amplify the nucleic acid. The amplified nucleic acid strand is detected with an oligonucleotide probe which specifically hybridizes to the amplified strand. Nucleic acid of chronic myelogenous leukemia

patient and that resulting from bcr-abl translocation were detected by the method.

AN 2000:85055 HCAPLUS <<LOGINID::20081125>>
DN 132:147583
TI Methods for detecting and measuring spliced nucleic acids and method of cytoplasmic nucleic acid preparation
IN Harvey, Richard C.; Eastman, Paul S.
PA Gen-Probe Incorporated, USA
SO PCT Int. Appl., 52 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 2000005418	A1	20000203	WO 1999-US16832	19990723 <--
	W: AU, CA, JP				
	RW: AT, BE, CH, DE, DK, ES, FR, GB, IT, LU, NL, SE				
	US 6849400	B1	20050201	US 1998-121239	19980723 <--
	CA 2337106	A1	20000203	CA 1999-2337106	19990723 <--
	AU 9951288	A	20000214	AU 1999-51288	19990723 <--
	AU 767568	B2	20031113		
	EP 1109932	A1	20010627	EP 1999-935912	19990723 <--
	EP 1109932	B1	20040616		
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
	JP 2002521037	T	20020716	JP 2000-561364	19990723 <--
	AT 269417	T	20040715	AT 1999-935912	19990723 <--
	ES 2221750	T3	20050101	ES 1999-935912	19990723 <--
PRAI	US 1998-121239	A	19980723	<--	
	US 1997-53509P	P	19970723	<--	
	WO 1999-US16832	W	19990723	<--	

RE.CNT 3 THERE ARE 3 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L7 ANSWER 11 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Methods for extracting RNA from biological samples
AB Disclosed is a safe and efficient method for extracting RNA from biol. samples, which method is comprised of (1) adsorbing RNA by mixing the sample with a neutral solution containing chaotropic substance and solid carriers (e.g. super magnetic metallic oxides); (2) washing the carriers with a low-salt (<100 mM) solution; (3) recovering RNA by heating the washed carriers. The neutral solution contains 4-7 M guanidine salt, 0-5% non-ionic surfactant, 0-0.2 mM EDTA, and 0-0.2 M reducing agents. The method was demonstrated by extracting RNA from Escherichia coli and hepatitis C virus RNA from a serum sample.
AN 1999:344509 HCAPLUS <<LOGINID::20081125>>
DN 131:40522
TI Methods for extracting RNA from biological samples
IN Sugiyama, Akio; Nishiya, Yoshiaki; Kawakami, Fumikiyo; Kawamura, Yoshihisa
PA Toyobo Co., Ltd., Japan
SO Jpn. Kokai Tokkyo Koho, 7 pp.
CODEN: JKXXAF
DT Patent
LA Japanese
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	JP 11146783	A	19990602	JP 1997-315295	19971117 <--
	JP 3812696	B2	20060823		

PRAI JP 1997-315295 19971117 <--

L7 ANSWER 12 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Rapid RNA and mRNA isolation procedure in the presence of a transition metal ion and oligo-dT-coated microparticles
AB Provided herein is a rapid method for isolating total RNA from test samples and further for isolating mRNA from test samples. The test sample is treated with a transitional metal ion having a valence of ≥+2 to form a precipitant and a supernatant. The supernatant is collect which should contain a purified solution of RNA. In cases where the nucleic acids are contained within organisms such as virus particles or cells, the test sample is treated with a lytic agent prior (a chaotropic agent, a salt, or a detergent such as RNAZol) prior to the transition metal ion treatment. Suitable ions include Co+3, Co+2, Zn+2, Cu+2, V+2, and Ni+2 in concns. >5 mM. The supernatant separation step includes passing the supernatant through a column with a solid-phase oligo dT matrix. The method is exemplified by the isolation of prostate-specific antigen mRNA from whole blood using RNAZol and a second step with oligo (dT)25-coated microparticles.
AN 1998:650071 HCAPLUS <<LOGINID::20081125>>
DN 129:272676
OREF 129:55529a,55532a
TI Rapid RNA and mRNA isolation procedure in the presence of a transition metal ion and oligo-dT-coated microparticles
IN Gundling, Gerard J.
PA Abbott Laboratories, USA
SO U.S., 6 pp.
CODEN: USXXAM
DT Patent
LA English
FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI US 5817798	A	19981006	US 1997-931981	19970917 <--
CA 2303398	A1	19990325	CA 1998-2303398	19980911 <--
WO 9914224	A1	19990325	WO 1998-US19043	19980911 <--
W: CA, JP				
RW: AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
EP 1017705	A1	20000712	EP 1998-944865	19980911 <--
EP 1017705	B1	20030806		
R: AT, BE, CH, DE, ES, FR, GB, IT, LI, NL				
JP 2001516763	T	20011002	JP 2000-511773	19980911 <--
AT 246698	T	20030815	AT 1998-944865	19980911 <--
ES 2205545	T3	20040501	ES 1998-944865	19980911 <--
PRAI US 1997-931981	A	19970917 <--		
WO 1998-US19043	W	19980911 <--		

RE.CNT 18 THERE ARE 18 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L7 ANSWER 13 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Method for enhancing chemiluminescence
AB The invention relates to a method for obtaining increased enhancement of luminescence from art known luminescent systems by the incorporation into the art known luminescent system of one or more detergents and one or more enhancer. Such enhanced luminescence can occur in solution or on a solid surface. The method can be practiced using anionic, cationic, zwitterionic, and non-ionic surface active or detergent compds. The method has broad application in any area where a signal generation system is required. Such areas include medical, veterinary, agricultural, and industrial diagnostics and quality control. This

includes any assay type designed to detect and/or quantitate the presence of any analyte, including industrial and pharmaceutical compds. as well as biol. compds. and organisms of all types such as proteins, carbohydrates, lipids, nucleic acids, bacteria and viruses. Examples of such tests include those utilizing nucleic acid probes, as well as immuno- and receptor-assays.

AN 1997:240626 HCAPLUS <<LOGINID::20081125>>
DN 126:222603
OREF 126:42987a,42990a
TI Method for enhancing chemiluminescence
IN Kohne, David E.
PA Kohne, David E., USA
SO PCT Int. Appl., 92 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI WO 9705209	A1	19970213	WO 1996-US12300	19960726 <--
W: AU, BR, CA, CN, FI, JP, KR, NO, NZ				
RW: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
AU 9666003	A	19970226	AU 1996-66003	19960726 <--
PRAI US 1995-1641P	P	19950728	<--	
WO 1996-US12300	W	19960726	<--	

L7 ANSWER 14 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Method for immobilizing nucleic acid molecules to be used in nucleic acid analysis
AB Synthetic nucleic acid mols. are non-covalently immobilized in the presence of a salt or cationic detergent on a hydrophilic polystyrene solid support containing an -OH, -C=O, or -COOH hydrophilic group or on a glass solid support. The support is contacted with a solution having a pH of about 6 to about 8 containing the synthetic nucleic acid and the cationic detergent or salt. Preferably, the cationic detergent is 1-ethyl-3-(3'-dimethylaminopropyl)-1,3-carbodiimide hydrochloride at a concentration of about 30 mM to about 100 mM or octyldimethylamine hydrochloride at a concentration of about 50 mM to about 150 mM. The salt is preferably NaCl at a concentration of about 50 mM to about 250 mM. When the detergent is 1-ethyl-3-(3'-dimethylaminopropyl)-1,3-carbodiimide hydrochloride, the glass support or the hydrophilic polystyrene support is used. When NaCl or octyldimethylamine hydrochloride is used, the support is the hydrophilic polystyrene. After immobilization, the support containing the immobilized nucleic acid may be washed with an aqueous solution containing a non-ionic detergent. The immobilized nucleic acid may be used in nucleic acid hybridization assays, nucleic acid sequencing and in anal. of genomic polymorphisms.
AN 1997:204423 HCAPLUS <<LOGINID::20081125>>
DN 126:261266
OREF 126:50524h,50525a
TI Method for immobilizing nucleic acid molecules to be used in nucleic acid analysis
IN Nikiforov, Theo; Knapp, Michael R.
PA Molecular Tool, Inc., USA
SO U.S., 25 pp., Cont.-in-part of U.S. Ser. No. 162,397, abandoned.
CODEN: USXXAM
DT Patent
LA English

FAN.CNT 9

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 5610287	A	19970311	US 1994-341148	19941116 <--
	CA 2155634	A1	19950615	CA 1994-2155634	19941206 <--
	WO 9515970	A1	19950615	WO 1994-US14096	19941206 <--
	W: AU, CA, JP				
	RW: AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
	AU 9513032	A	19950627	AU 1995-13032	19941206 <--
	AU 682741	B2	19971016		
	EP 684952	A1	19951206	EP 1995-904282	19941206 <--
	EP 684952	B1	20041013		
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LI, LU, MC, NL, PT, SE				
	AT 279427	T	20041015	AT 1995-904282	19941206 <--
PRAI	US 1993-162397	B2	19931206	<--	
	US 1994-341148	A	19941116	<--	
	WO 1994-US14096	W	19941206	<--	

L7 ANSWER 15 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN

TI Isolation of nucleic acid from biological sample, method comprising nucleic acid binding to solid support then separation from support, and kit comprising detergents and other components

AB The present invention provides a method of isolating nucleic acid from a sample, said method comprising contacting said sample with a detergent and a solid support, whereby soluble nucleic acid in said sample is bound to the support, and separating said support with bound nucleic acid from the sample. Where the method of the invention is used to isolate DNA, it may conveniently be coupled with a further step to isolate RNA from the same sample.

AN 1996:458048 HCAPLUS <<LOGINID::20081125>>

DN 125:107039

OREF 125:19863a,19866a

TI Isolation of nucleic acid from biological sample, method comprising nucleic acid binding to solid support then separation from support, and kit comprising detergents and other components

IN Deggerdal, Arne Helge; Larsen, Frank

PA Dynal A/s, Norway; Dzieglewska, Hanna Eva

SO PCT Int. Appl., 53 pp.

CODEN: PIXXD2

DT Patent

LA English

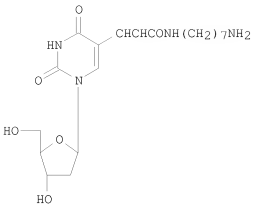
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 9618731	A2	19960620	WO 1995-GB2893	19951212 <--
	WO 9618731	A3	19960912		
	W: AL, AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK				
	RW: KE, LS, MW, SD, SZ, UG, AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG				
	CA 2207608	A1	19960620	CA 1995-2207608	19951212 <--
	AU 9641829	A	19960703	AU 1996-41829	19951212 <--
	AU 706211	B2	19990610		
	EP 796327	A2	19970924	EP 1995-940351	19951212 <--
	EP 796327	B1	20040728		

```
        R:  AT, BE, CH, DE, FR, GB, IT, LI, SE
     JP 11501504          T     19990209    JP 1996-518463          19951212 <--
     JP 3787354           B2    20060621
     AT 272110            T     20040815    AT 1995-940351          19951212 <--
     US 20040215011       A1    20041028    US 1997-849686          19970821 <--
     US 20060058519       A1    20060316    US 2005-234001          20050923 <--
     US 7173124           B2    20070206
     US 20070190559       A1    20070816    US 2007-671426          20070205 <--
PRAI GB 1994-25138        A     19941212  <--
     WO 1995-GB2893       W     19951212  <--
     US 1997-849686       A1    19970821  <--
     US 2005-234001       A1    20050923

L7   ANSWER 16 OF 21  HCAPLUS  COPYRIGHT 2008 ACS on STN
TI   Immobilization of nucleic acids on solid supports such as glass
     and polystyrene using salt and cationic detergent
AB   An improved method for immobilizing a synthetic nucleic
     acid mol., such as an oligonucleotide, to a solid
     surface, especially polystyrene or glass is described.  The method comprises
     incubating the nucleic acid with the solid
     support in the presence of a salt or cationic detergent
     , then washing with an aqueous solution  The method is useful in facilitating
     polymorphic analyses, in hybridization assays, and in solid
     -phase DNA sequencing.  Immobilization of oligonucleotide probes on
     polystyrene using NaCl and 1-ethyl-3-(3'-dimethylaminopropyl)-1,3-
     carbodiimide hydrochloride followed by washing with a buffered solution
     containing Tween 20 was described.  The immobilized probes were used
     for detection of PCR products.
AN   1995:789412  HCAPLUS <<LOGINID::20081125>>
DN   123:190499
OREF 123:33713a,33716a
TI   Immobilization of nucleic acids on solid supports such as glass
     and polystyrene using salt and cationic detergent
IN   Nikiforov, Theo; Knapp, Michael R.
PA   Molecular Tool, Inc., USA
SO   PCT Int. Appl., 61 pp.
     CODEN: PIXXD2
DT   Patent
LA   English
FAN.CNT 9
     PATENT NO.          KIND   DATE        APPLICATION NO.         DATE
     --------------      ----   --------    -------------------     --------
PI   WO 9515970          A1     19950615    WO 1994-US14096         19941206 <--
         W:  AU, CA, JP
         RW: AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE
     US 5610287           A      19970311    US 1994-341148          19941116 <--
     AU 9513032           A      19950627    AU 1995-13032           19941206 <--
     AU 682741            B2     19971016
     EP 684952            A1     19951206    EP 1995-904282          19941206 <--
     EP 684952            B1     20041013
         R:  AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LI, LU, MC, NL, PT, SE
     AT 279427            T      20041015    AT 1995-904282          19941206 <--
PRAI US 1993-162397       A      19931206  <--
     US 1994-341148       A      19941116  <--
     WO 1994-US14096      W      19941206  <--

L7   ANSWER 17 OF 21  HCAPLUS  COPYRIGHT 2008 ACS on STN
TI   Method for immobilization of polynucleotide (gene probe)
GI
```



II

AB Immobilization of a polynucleotide involves linking a polynucleotide containing a nucleotide having at least one linker arm to a solid -phase support, wherein the linker arm G-P-Q (G = residue derived by removing the position 1-C atom of the reducing terminus from ribose or deoxyribose; P = adenosine, guanosine, cytidine, thymidine, uridine; Q = C1-15 monovalent organic group). A method for detecting a target nucleic acid involves immobilization of a polynucleotide according to the above method, hybridization of a sample containing the target nucleic acid, subsequent hybridization of a labeled probe, and detection of the label. This immobilization method improves the efficiency for immobilization of a gene probe (polynucleotide) on a solid support and enables detection of a target nucleic acid by hybridization method with high sensitivity. Thus, an oligonucleotide capture probe 5'-CGGTCATTCTGCTGXGTTCGTAAAAT-3' (I; X = linker arm-containing nucleoside represented by II) and a DNA probe 5'-CCCCGGTTCTGAXGAGATATTGTT-3' (III; X = same as above) were prepared by a DNA synthesizer using the phosphoramidite method. The probe III was dissolved in 0.2 M aqueous NaHCO3, reacted with suberic acid disuccinimidyl ester, purified by a gel filtration column of Sephadex G-25, and then condensed with alkali phosphatase to give alkali phosphatase-labeled probe (IV). The capture probe I containing II was diluted with 50 mM phosphate buffer to 10 pmol/mL and the diluted solution (100 mL) was added to each well of a polystyrene microtiter plate and incubated at room temperature followed by removing the capture probe solution with an aspirator, adding a block buffer (150 μL), and incubating for blocking at room temperature for 2 h. After removing the block buffer from the wells, a sample solution (10 μL) of enteritis vibrio genes (denatured by 0.6 N aqueous NaOH) and subsequently a hybridization buffer were added and hybridized at 50° for 60 min. The liquid was removed and the wells were successively rinsed with a rinsing solution (2 + SSC, 10% sodium lauyrylsulfate) (200 μL) at 50° for 60 min and a rinsing solution (1 + SSC, 200 μL) followed by adding a solution of the labeled probe IV, incubating it at 50° for 60 min, removing the probe solution from wells, successively rinsing the wells with a rinsing solution (1 + Steady-State Creep, 0.5% Triton X-100) (200 μL) at 50° for 60 min and a rinsing solution (1 + SSC, 200 μL). A solution of Lumiphos 480 (chemiluminescence substrate for alkali phosphatase) (100 μL) was added for the luminescence reaction which was carried out at 37° in dark for 15 min. The detected luminescence was 7.244 and 0.010 for the pos. and neg. enteritis vibrio gene samples, resp., vs. 2.902 and 0.013 when a capture probe without the arm linker was used. The arm linker-containing capture probe I increased the signal to noise (S/N) ratio from 223.2 to 724.4.

AN 1995:358864 HCAPLUS <<LOGINID::20081125>>
DN 122:182725
OREF 122:33376h,33377a
TI Method for immobilization of polynucleotide (gene probe)
IN Daimon, Katsuya; Yoshimoto, Misao; Hayashi, Satoko
PA Toyo Boseki, Japan; Toyobo Co., Ltd.
SO Jpn. Kokai Tokkyo Koho, 8 pp.
CODEN: JKXXAF
DT Patent
LA Japanese
FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI JP 06329694	A	19941129	JP 1993-118615	19930520 <--
JP 3596620	B2	20041202		
PRAI JP 1993-118615		19930520	<--	

L7 ANSWER 18 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Rapid screening of recombinant plasmids with a non-radioisotopic hybridization assay
AB Here the application of a non-radioisotopic hybridization assay, the DNA enzyme immunoassay (DEIA), to the screening of recombinant plasmids is reported. The DEIA assay, which was originally proposed for the detection of amplification products, is based on the use of a monoclonal antibody (MAb) that specifically reacts with double-stranded DNA. The particular characteristics of the anti-DNA MAb, now com. available (Sorin Biomedica, Saluggia, Italy), allow the revelation of the hybridization event between any cDNA sequences, without limitations in nucleotide composition or probe length. Briefly, a specific oligonucleotide probe, fixed on a solid support by an avidin-biotin bridge, was hybridized with denatured plasmid DNA. The probe, modified at the 5' end by the introduction of a primary amino group, was synthesized by the Model 391 PCR-Mate EP DNA synthesizer. Biotinylation of 5'-modified oligonucleotide was performed as previously described. Streptavidin-coated microplates were incubated overnight at 4° with 10-100 ng of the biotinylated probe in 100 μL of TE buffer. The solid phase was then washed four times, just before use, with 200 μL of washing solution (6.7 mM phosphate buffer, pH 6.4 0.13M sodium chloride, 0.004% Cialit [2-ethylmercurithio-5-benzoxal-carboxylic acid, sodium salt], 0.1% Tween 20). Hybridization with the denatured plasmid DNA was revealed by the use of the anti-DNA MAb and of a peroxidase-labeled anti-mouse Ig antibody. The result was read at the spectrophotometer at 450 nm.
AN 1993:161860 HCAPLUS <<LOGINID::20081125>>
DN 118:161860
OREF 118:27577a,27580a
TI Rapid screening of recombinant plasmids with a non-radioisotopic hybridization assay
AU Ravaggi, A.; Mantero, G.; Albertini, A.; Primi, D.; Cariani, E.
CS Sch. Med., Univ. Brescia, Brescia, 25123, Italy
SO BioTechniques (1992), 13(4), 506, 508
CODEN: BTNQDO; ISSN: 0736-6205
DT Journal
LA English

L7 ANSWER 19 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI High salt lysates: a simple method to store blood samples without refrigeration for subsequent use with DNA probes
AB Blood specimens to be tested for the presence of Plasmodium falciparum using specific DNA probes can be stored as high salt lysates (HSL) without refrigeration. The lysates are prepared from 100 μL blood

samples by a simple 3-step procedure using 2 vols. of H2O to lyse the erythrocytes (step I), 1 volume of a detergent/EDTA mix to lyse the parasites (step II), followed by the addition of 1 volume cesium trifluoroacetate (step III). The parasite DNA was found to be undegraded, as shown by the unaltered pattern of repetitive sequences obtained after storage of up to 1 mo at 37°, due to the inhibition of DNA degrading enzymes by the cesium salt. The bulk of protein can be removed from the samples by a 1-step precipitation The addition of 0.3 vols. of a mixture of ethanol:chloroform:isoamyl alc. (2.5:1:0.04 volume/volume) ppts. >90% of the proteins from the lysates, leaving >86% of the parasite DNA in the supernatant. The reduced protein content of the samples, when applied to solid supports, results in an increased signal:background ratio on autoradiograms.

AN 1988:566575 HCAPLUS <<LOGINID::20081125>>
DN 109:166575
OREF 109:27551a,27554a
TI High salt lysates: a simple method to store blood samples without refrigeration for subsequent use with DNA probes
AU Zolg, J. Werner; Scott, Ethel D.; Wendlinger, Monika
CS Dep. Mol. Biol., Biomed. Res. Inst., Rockville, MD, 20852, USA
SO American Journal of Tropical Medicine and Hygiene (1988), 39(1), 33-40
CODEN: AJTHAB; ISSN: 0002-9637
DT Journal
LA English

L7 ANSWER 20 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Assay utilizing ATP encapsulated within liposome particles
AB An assay for an analyte (e.g. antigen, DNA probe) of a specific binding pair (ligand, antiligand) comprises (a) combining a fluid sample with a solid support sensitized with receptors for the analyte; (b) contacting the support with ATP-loaded liposomes having bound ligand, ligand analog, or antiligand; and (c) testing for the presence of ATP associated with the support. ATP released from the liposomes is detected by a luciferin-luciferase reagent and a luminometer. ATP-loaded liposomes were prepared from L-α-dipalmitoylphosphatidylcholine, N-3-(2-pyridyldithiopropionyl)dipalmitoylphosphatidylethanolamine, cholesterol, and ATP in CHCl3, Et2O, and MeOH. Fab' fragments of anti-Group A Streptococcus antibody was reacted with the liposomes which were then used in a rapid immunoassay for Group A Streptococcus. Anti-Group A Streptococcus-coupled polystyrene particles were reacted with Group A Streptococcus extract and anti-Group A liposomes for 20 min; then the reactants were centrifuged and washed with phosphate-buffered saline. Triton buffer, luciferin-luciferase, and releasing agent were added, and light emitted was read in a luminometer.
AN 1988:109228 HCAPLUS <<LOGINID::20081125>>
DN 108:109228
OREF 108:17827a,17830a
TI Assay utilizing ATP encapsulated within liposome particles
IN Bernstein, David
PA New Horizons Diagnostics Corp., USA
SO U.S., 7 pp.
CODEN: USXXAM
DT Patent
LA English
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 4704355	A	19871103	US 1985-716702	19850327 <--

PRAI US 1985-716702 19850327 <--

L7 ANSWER 21 OF 21 HCAPLUS COPYRIGHT 2008 ACS on STN
TI Interactions of proteins and nucleic acid
AB Both the nucleus and the protoplasm of the red cells of Triton are precipitated by sulfosalicylic acid, whereas only the nucleus is precipitated by La salts (photomicrographs). The nuclei precipitated first, before precipitation of the protoplasm began. At a certain stage, cell nuclei are entirely protein-free. Nucleic acid can be precipitated as the La complex from a solution containing also protein by using a mixture of La salt + malonic acid, the latter in high concentration The protein stays in solution as the strong affinity of nucleic acid for protein is blocked by malonic acid. By applying formalin + Zn ions to a protein nucleic acid solution, protein is precipitated and nucleic acid remains in solution When a solid mixture of proteins and Na salt of nucleic acid is treated with a trypsin preparation containing La ions, the protein goes into solution (except histone which is not quantitatively removed) and nucleic acid remains undissolved. By using the enzyme-La or the La-malonic acid reagents it should be possible to detect protein structures in the cells by photographing at such a wave length that the proteins absorb a sufficient amount of light. Application of this method to analysis of chromosomes from testicular cells results in the finding that proteins were not packed as solid proteins at intervals in the chromosomes. The solubility of the La salt of nucleic acid (thymus) is very small (less than 0.001% at a La concentration of 10-4 mole/1.). The decomposition of nucleic acid by the La-malonic acid reagent is negligible in the course of the first hr. only. Stenobotrus cells were digested with the trypsin-La reagent. The chromosomes were rendered beautifully visible, in ordinary light, when mitotic cells were digested.
AN 1935:61094 HCAPLUS <<LOGINID::20081125>>
DN 29:61094
OREF 29:8031i,8032a-d
TI Interactions of proteins and nucleic acid
AU Caspersson, T.; Hammarsten, E.; Hammarsten, H.
SO Transactions of the Faraday Society (1935), 31, 367-89
CODEN: TFSOA4; ISSN: 0014-7672
DT Journal
LA Unavailable